Review Article

Studies of Aniline and it's Derivatives on Various Organ Systems: A Mini Review

Nayan Kumbhare^{1*(D)}, Varsha Dhurvey ^{2(D)}

¹Department of Zoology, RTM Nagpur University, Nagpur, Maharashtra, India ²Department, of Zoology, RTM Nagpur University, Nagpur, Maharashtra, India

*Corresponding Author: 🖂

Received: 17/Mar/2025; Accepted: 04/Apr/2025; Published: 30/Apr/2025. | DOI: https://doi.org/10.26438/ijsrbs.v12i2.672

Copyright © 2025 by author(s). This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 International License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited & its authors credited.

Abstract- Certain chemical pollutants in the environment can change a variety of biological processes. Anilines are widely used in the manufacturing of plants, colours, medications, pesticides, and herbicides. This review aims to highlight the destructive, dangerous and altering effects of aniline on various organisms. We believe that this study will provide helpful information regarding aniline's toxicological research, which will help in the formulation of acceptable guidelines for aniline usage.

Keywords- Aniline, reproductive toxicity, neurotoxicity, nephrotoxicity, carcinogens

1. Introduction

Numerous industries, including electronics, pharmaceuticals, agriculture, and many more, have benefited greatly from human contributions. The development of industries depends on chemicals. The features and attributes of a material are often what define its usage [1]. Aniline, the main molecule of the family of aromatic amine, is a crucial component of many of these materials. Aniline or its derivatives, including 3,4dichloroaniline, 4,4'-methylenedianiline, and 1naphthylamine, are precursors or intermediates in a wide range of rubbers, medications, insecticides, dyes, cosmetics, and adhesive goods [2,3,4,5]. Several anilines have been identified as high-volume production chemicals by a number of countries, including the US and the EU [6,7,8]. In 2016, an estimated 5.6 million tons of aniline were produced globally [5].

Ammonia, phenol, and benzene can all be used to make aniline in an industrial setting. It smells less like rotten fish and more like volatile amines. It has the property of aromatic chemicals and burns with a smoky flame right away. According to [1] this colourless, hydrophilic molecule can also mix with various organic solvents. The production of precursors for the polymer polyurethane is its primary application.

Considered to be among the best thermal insulators, it is utilized in every refrigerator. Additionally, it is used as a pesticide in addition to agricultural chemicals. It starts with the pigments and colours found in aniline. It is utilized in both medicinal and photographic chemicals. Paracetamol, a well-known analgesic drug, is made from aniline. A dye known as O-toluidine is used in forensic science to extract suspected blood.

Anilines are employed as precursors in manufacture of numerous colours and some care articles and could be extracted from a wide range of materials. For example, 1-NPA can be freed from the hairs colour, tattoos, mehendi and make up products [9]. While residual isocyanates in polyurethane adhesive can generate 4,4'-MDA, rubber products can emit 1-NPA and 4,4'-MDA at certain temperatures [10]. Pharmaceuticals may also be a supply of aniline in the environment because aniline is used to create pain killers, fever reducing, allergy controlling, and multivitamins [11]. As breakdown products, these products may be exposed to the environment.

For instance, 1-naphthylamine from inks, pesticides, and herbicides (Guzman Mar *et al.*, 2006), 3,4-DCA from pesticides such as diuron, propanil and linuron [4,12,13], and 4,4'-methylenedianiline from rubber and polyurethane compounds [14].

Some anilines are commonly discovered in sediments and rivers around the world, despite the lack of information on their environmental quantities [15,16,4]. Levels of 3,4-DCA



as high as 68.2ng/L were detected in water from the Chattahoochee River and Scope Creek in the United States [17]. 1. In the Zonguldak industrial area of Turkey, NPA was detected in river sediment at an absorption of 186.45 ng/kg [18]. 3,4-dichloroaniline's precursor, diuron, was detected at concentrations as high as 230 ng/L in Korea's main harbours and as high as 1360 ng/L in a port in Jinhae Bay [19].

Studies have been conducted on the toxicological consequences of several anilines due to their diverse sources and environmental detection. The review's objective is to raise awareness of aniline's hazardous and detrimental effects on animal test models. We believe that by providing valuable information regarding aniline's toxicological profile, this analysis will aid in the development of safe guidelines for its appropriate usage.

Table-1	Aniline's	and its	derivatives'	characteristics:

Name	Structure	Molar mass (g/mol)	Reference
Aniline	SHN-	93.13	(National Institute of Health)
3,4- dichloroaniline	H ² CI	162.013	(National Institute of Health)
1- Naphthylamine	NH ₂	143.19	Bhuiyan <i>et al.</i> , 2019
4-chloroaniline	Ĩ Z - Ū	127.57	Bhuiyan <i>et al.</i> , 2019
4,4'-methylene dianiline	H ₂ N NH ₂	198.269	Bhuiyan <i>et al.</i> , 2019

2. Materials and Methods

Literature survey strategy

The information collected by google scholar, Meta-Analyses, or PRISMA by the writer of the paper. Most of the time, both meta-analyses and systematic reviews are brought out using approval. Recommendations are a small set of evidence-based suggestions. The author used three data platforms - Science Direct Google Scholar, PubMed, and -to concentrate regarding both non-experimental and experimental research. Search engines were put to use to find research information published from 1971 and 2025.

Data extraction

The studies that appeared in the search engines were the first to be scanned. Relevant studies were imported into Microsoft Excel and totalled according to their abstract, purpose, and conclusions.

3. Results and Discussion

Studies of aniline on various organ systems:

3.1. Study of aniline on nervous system: [20] Investigated aniline's neurotoxicity and age-dependent effects in male rats. Two of the six rats that were given 1,000 mg/kg of aniline at the age of 4 weeks, exhibited symptoms paralysis of the hind limb or gait which was parallactic in middle of after treatment days 15 and 8. On day 15 following treatment, the white matter of spinal cord in the rats which were administered 750 or 1,000 mg/kg aniline at the age of 4 weeks, displayed spongy change. In addition to mild peripheral nerve degeneration, 3 of the 6 rats given 1000 mg/kg body weight aniline showed some changes revealed alterations in the medulla oblongata and pons spinal trigeminal tracts and facial nerve. [21] Studied that exposure to aniline particularly results in neuron poisoning and a range of neurological symptoms, such as sarcoma, that is typified by fibrosis, splenomegaly, and the formation of tumours. According to the literature that is available, aniline has a neurotoxic effect.

Table-2 Study	of aniline on	nervous system
---------------	---------------	----------------

Animal model	Aniline dose	Time period	Inference	Reference
Rats	500, 750 or 1,000 mg/kg	15 days	Signe of paralytic gait or hindlimb	Okazaki <i>et al.</i> ,
			paralysis.	2001
Rats	-	-	Weakness, lethargy, ataxia, vertigo,	Makhdoumi et
			nausea, vomiting, and tinnitus fatigue.	al., 2019

3.2 Study of aniline on nephric system- In a study by [22], the nephrotoxic potential of aniline was assessed in vitro and in vivo using Fischer 344 rats. Renal function was evaluated at 48 and 24 hours after rats in the treated group received aniline (1.0, 1.5, or 0.4 mmol/) and rats in the control group received an intraperitoneal injection of saline (2.0 ml/kg). 2. Aniline decreased urine production, raised blood urea nitrogen levels, and prevented lactate and basal-stimulated p-aminohippurate collection by renal cortical at a dosage of 1.0 mmol/kg. At the dosages used in this study, aniline had no discernible effect on renal function. Aniline (10–4 M or above) decreased the base-level buildup of p-aminohippurate in the in vitro assays.

In a study by [23], male Fischer 344 rats were given intraperitoneal injections of aniline, acetyl-p-aminophenol, pnitroaniline, p-aminophenol, p-chloroaniline, pchloronitrobenzene, or p-anisidine at a dosage of 1.0 mmol/kg. Rats with p-aminophenol injections showed necrosis of renal tubular epithelial cells and a significant increase in urine and γ -glutamyl transpeptidase and N-acetyl- β -D-glucosaminidase functioning. The tubular epithelial cells in rats responded to p-nitroaniline and p-anisidine by swelling. However, the administration of aniline or its derivatives had no effect on the urine enzymes or renal histology. The toxicity of the renal cortical slices was assessed by tracking changes in tissue gluconeogenesis capability and lactate dehydrogenase release. None of the anilines enhanced the release of lactate dehydrogenase. 4-iodoaniline 2.0 mM caused the greatest reduction in gluconeogenesis ($92\%\downarrow$), while 4-bromoaniline decreased gluconeogenesis at a dose of (0.1 mM). The most powerful nephric toxicant was 3,5-dibromoaniline, while the least powerful was 3,5-difluoroaniline [24].

A different study was performed in which male Wistar rat's male were used to identify and assess the nephrotoxic hazards of aniline. Group A rats were dosed with 20 mg/kg body weight aniline for a period of 30 days, while B group rats were treated with regular saline water. Blood urea, creatinine, and a few serum pointers of kidney harm all increased in concentration. Other than histological changes such as the shedding of the brush edge of the proximal tubular cells, sodium and potassium contents dropped. Significant degenerative alterations were seen in the distal tubular cells and proximal tubular cells [25].

Lable-5 Study of annue on nephile system

			-	_
Animal model	Aniline dose	Time period	Inference	Reference
Fischer 344	(0.4, 1.0, or 1.5	-	Decreased urine production, elevated blood	Rankin et al., 1986
rats	mmol/kg),		urea nitrogen levels, and prevented renal	
			cortical slices from accumulating p-	
			aminohippurate (PAH) in response to lactate	
			and baseline stimuli.	
Male Fischer	1.0 mmol/kg	-	Necrosis of renal tubular epithelial cells and	Yoshida et al.,1989.
344 rats			a considerable increase in the activity of γ -	
			glutamyltranspeptidase (γ-GTP) and N-	
			acetyl-β-D-glucosaminidase (NAG) in the	
			urine were observed.	
-	-	-	Reduction in gluconeogenesis (92%↓), while	Hong et al., 2000.
			4- bromoaniline decreased	
			gluconeogenesis in the dose of 0.1 mM	
Male Wistar rats	20 mg/kg	30 days	Creatinine and blood urea and serum	Ramteke and
		-	indicators of kidney damage all rose in	Dhurvey 2020.
			concentration	

3.3 Study of aniline on hepatic system- [26] conducted a study in which rats were used to examine the effects of administering aniline metabolites, specifically phenylhydroxylamine and p-aminophenol, and human volunteers were utilized to examine the effects of oral aniline administration. Rats fed phenylhydroxylamine or aniline developed splenic enlargement, hemoglobinemia, and Heinz bodies. An oral dose of aniline (20 mg/kg body weight) showed no effect on rats. Five or fifteen milligrams of aniline taken orally did not cause any reaction in twenty human

participants. Doses ranging from 25 to 65 mg significantly increased haemoglobin levels in the blood, although no Heinz bodies were observed.

A study found that the liver hydroxylates aniline to make a number of derivatives, few of them, end up in RBCs. The aniline underwent biotransformation in the livers of male albino rats in the concentration of 50mg/kg BW [27].

In a study by [28] mice received dose of 120, 60, 30, 15, 7.5 and 0 mg/kg BW of p-Chloroaniline for 13 weeks, whereas rats given oral dosage of 5, 40, 20, 10, 80 and 0 mg/kg BW of p-Chloroaniline. In mice and rats, the treated animals' bodyweight decreased. Methemoglobin levels in all of the dosed mice and rats were higher than that in the control.

The male mice's livers showed a collection of lymphocyte buildup, inflammatory cells, large Kupffer cells, proliferation in the bile ducts, and portal vein dilatation after taking 1 to 8th of 3,4DC-A(dichloroaniline) for 35, 29, and 14 noes of day. [29].

Aniline was administered to the male albino rat's reared hepatocytes for a full day at doses of 10, 5.0, 50, 2, 2.5, 1.25, and 0 g/mL. In contrast to the control, aniline significantly decreased catalase, superoxide dismutase, glutathione, and the membrane potential of mitochondria while increasing malondialdehyde and reactive oxygen species in hepatocytes. Aniline reduced cell life and cell death was seen in a dose-dependent fashion. [5].

The liver weight of huge blue F344 (male) rats increased statistically significantly when given a dose of hundred per kg aniline or fifteen mg per kg para-chloroaniline. Additionally, the rats' calcium and cholesterol levels were higher than those of control animals [30].

Furthermore, the addition of aniline caused spectrum aberrations in liver microsomes, oxidative damage from aniline, and hepatocyte death. Aniline exposure over an more duration of time in vivo results in a toxic response in the spleen that includes fibrosis, hyperplasia, tumor growth, and splenomegaly. Due to oxidative DNA damage, iron overload, and accelerated cell proliferation caused by aniline, the spleen may show malignancies [31].

Animal model	Aniline dose	Time	Inference	Reference
		period		
Rats	5 mg and 15 mg	-	Rise in hemoglobin	Jenkins et al.,
				1972.
Rats	50 mg/kg body	-	The liver hydroxylates aniline to create a variety of	Eyer et al.,
	weight		derivatives, few of them become lodged in red -blood	1980.
			cells.	
Rats	0, 7.5, 15, 30, 60,	13 weeks	All dosed rats and mice had considerably higher	Chhabra <i>et</i>
	and 120 mg/kg BW		haemoglobin levels than the control group.	al.,1991.
Male Swiss	-	14, 30, and	Showed accumulation of infiltrating lymphocytes,	Sharma et al.,
albino mice		35 days	large Kupffer cells,	2008.
-	-	-	Aniline induced apoptosis and reduced cell viability	Wang et al.,
			in relation to concentration	2016.
Big Blue F344	15 mg/kg	-	Liver weight increased statistically considerably,	Koenig et al.,
(male) rats'			and their calcium and cholesterol levels were	2018.
			significantly higher	
-	-	-	Aniline causes fibrosis, hyperplasia, tumor growth,	Mohurle et
			and splenomegaly. DNA damage, and accelerated cell	al., 2023.
			growth.	

Table-4 Study of aniline on hepatic system

3.4 Study of aniline on reproductive system- According to a study done by [32] the rats which were given aniline caused a marked decrease in sterol 3-beta dehydrogenase and many lipids storing clear cells in their corpora lutea. The endoplasmic reticulum shrank or vanished entirely, indicating a significant reduction in the number of enzymes that perform steroidogenesis. This shows aniline disrupts the corpora lute a's steroidogenesis. Therefore, it may be concluded that aniline changes the rat steroidogenesis.

Accordingly, Fischer 344 rats that were pregnant were dosed with aniline hydrochloride (30, 100 or 10 mg/kg/day) from gestational day 7 to birth in an experiment by [33]. Rats were

seen not teratogenic to aniline hydrochloride. Therefore, it may be said that treatment with aniline is not found to be teratogenic at low dosages. In a work of [34] the reproductive toxicity of p-nitroaniline in rats of the F2, F1, and F0 generations was studied. The First generation showed no change in either the male fertility index or the pregnancy rate. Histopathological analysis of the F0 animals' epididymides and testes showed no such alterations that might affect the reproductive capacity. Therefore, this concludes that less aniline levels have no effect on mating, lactation, or gestation.

[35] Conducted an experiment that showed the effect of 3-4 dichloroaniline on histology of gonads in the estuary Mysid

Mesopodopsis slabberi. For a period of 48 hrs, the mysids were exposed to semi lethal concentrations of 3,4-DCA (1.40, 0.10, 1.00 0.30, 0.50, 0.90, 1.20, 1.10, 1.30, and mg/L). Several tissues were examined histologically, and animals given doses more than 0.30 mg/L showed damage. Gonads were obviously impacted by 3,4-DCA, as seen by the buildup of this hazardous material and structural abnormalities. Therefore, it may be said that aniline derivatives may be harmful to fish reproductive health. Accordingly, [36] explained how 3,4-dichloroaniline affected the pointers of enzyme activity of testis in rats. Alkaline phosphatase and acid phosphatase activities in the study significantly enhanced at less 3,4-dichloroaniline dose and became less in higher 3,4-DCA concentration.

In a different study by [37] male Wistar rats were give aniline at dose of 13.33, 27.67,55.33 mg/kg body weight for a duration of 1 month. The result showed a decline in mitotic rate of the spermatocytes.

In another study by [38] female Wistar rats were given 93, 31 mg/kg BW aniline 7 days after intercourse to birth. There was a decline in the distance between the genitals and anus of the pups.

[39] Used H295R cell line and exposed them to 10, 100 mg/l BW aniline. There was an more expression of CYP 19A and a less expression of StAR or CYP 17 genes. The author

extended his study in the same year on male zebra fish which was exposed to 0. 024, 0.12, 3.0 0.6 mg/L of 3,4-Dichloroaniline; 5.0 ,0.04, 1.0, 0.2, or 5.0 mg/L of 1-Naphthylamine and 25, 5.0, 1.0, 0.2 mg/L for 4,4'-Methylenedianiline. Decline in the testosterone amount and higher 17-estradiol/testosterone ratios was seen in the results.

In a study by [40] the reproduction activity of the Oryzias (ricefish), fish surrounded to 3,4-DCA was investigated. The spawning rate and fertilization rate became very less in the group that was given to the highest concentration (250 gm/L). The gonad and somatic index GSI were found to be incredibly less in fishes that were put to 250 g/L dose. The outcome proves that 3,4-dichloroaniline hampers Oryzias (ricefish), reproduction by affecting gonadal tissue and fertility. Therefore, aniline reduces the Javanese medaka fish's reproductive capacity.

According to [41] the reproduction system is affected by aniline. It has shown to decrease reproductivity in fishes and affect the gonads' histology. It changes the number of hormones in rats, which impacts the animal's ability to conceive. It also results in chromosomal abnormalities and lowers male sperm counts.

Model animal	Aniline dose	Time period	Inference	Reference
Female rat	-	-	The corpus lutea displayed cells that	Hatakeyama
			stored lipids. Sterol 3-beta	<i>et al.</i> , 1971
			dehydrogenase decreased	
Female rat	30,10,100mg/kg bw	From	Toxicology in mothers	Price et al.,
		gestational day	and decline in the f0 generation	1985
		7 till birth	fertility ratio.	
Male and female rat	1.5, 9.0 0,0.25 in oral	14 weeks		Nair et
	corn oil	before to		al.,1990
		mating, during		
		mating,		
		gestation and		
		lactation		
Female and male Mysid	1.10, 1.20,	48 h	Damages the ovaries and histological	Sardo et al.,
Mesopodopsis slabberi	1.30,1.40, 0.10,0.30,0.50		structures	2005
fish	,0.90, mg/l			
Male rat	81, 39 mg/kg bw orally	-	At a low concentration of 3,4-	Zhang and
			dicholroaniline, the activities of	Lin, 2009
			(alkaline phosphatase) and ACP (acid	
			phosphatase) greatly rose, whereas at	
			a greater concentration, they	
			dramatically decreased.	
Male rat	27.67,55.33,13.33mg/kg	30 days	Reduction in the spermatocytes'	Eissa <i>et</i>
	BW orally		mitotic index	al.,2012
Female rat	93 ,31mg/kg BW	7 days post	Decrease in pups' AGD	Holm <i>et al</i> .,
		intercourse till		2016
		birth		
H295R cell line	100, 10mg/l BW	-	CYP19A up-regulation and StAR or	Bhuiyan et
			CYP17 gene down-regulation.	al., 2019

Toblo 5	Study	of aniling	on ron	roductivo	evetom
Table-5	Study	or annine	on rep	roductive	system

Japanese rice fish (Oryzias latipes)	250g/l	Slowing in the gonadal development	Ibrahim <i>et</i> <i>al.</i> , 2021
		It has shown to lessen the reproductivity in fishes and affect the	Ingle <i>et al.,</i> 2023
		gonads' histology. It has shown to lessen the reproductivity in fishes	
		and affect the ovaries and testis histology, this alters the hormonal	
		quantity in animal, that causes an effect on the fertility.	

3.5 Study of aniline on genes and cancer- According to a study by [43], no genes alteration, was observed in the silk worm locus test when oocyte at the pupa stage were given about $25\mu g$ of aniline.

Aniline hydrochloride was used to lung cells and Chinese hamster don cells. Without any chromosomal abnormalities, sister chromatid exchange was initiated. Aniline hydrochloride was thus found to have no clastogenic action [59]

In a study by [46], 4-chloroaniline was found to be genotoxic to the liver cells of mice, rats, and hamsters. DNA was repaired when MOCA was administered to hepatocytes from hamsters, mice, and rats. Rats were administered p-Chloroaniline deionized (PCA) water by gavage at rates of 6, 2, 0, or 18 mg/kg body weight for five days over a period of 103 weeks in an experiment by [47]. Rats of both sexes received doses of 10 and 30 mg/kg on a comparable schedule. All of the animals treated with PCA showed a marked rise in splenic fibrosis. Additionally, splenic sarcomas were observed.

Pheochromocytomas of the adrenal glands were slightly more common. Medication-treated mice had greater incidences of adenomas or hepatocellular carcinomas. Hepatic or splenic hemangiosarcoma's also increased in the high-dose group. Based on these results, we conclude that aniline causes cancer in both mice and rats. At high concentrations, it leads to the development of sarcomas and carcinomas, but at low dosages, it has no mutagenic effects. Aniline raises the risk of developing sarcomas and carcinomas, claims [41].

Male Sprague Dawley rats were given 300 mg/kg BW of aniline orally in a study by [44]. During the first 24 hours following the treatment, urine was collected. The results of the investigation showed that aniline was not mutagenic. However, urine metabolites of aniline were mutagenic. Male Swiss mice were given a single intraperitoneal injection of aniline at dosages ranging from 61 to 420 mg/kg BW.

The experiments' result proves that the amount of sister chromatids exchange in the bone marrow per metaphase rose [45].

Animal model	Aniline dose	Time period	Inference	Reference
Lung cells and Chinese hamster don cells	-	-	Chromosomal alterations were not seen	Abe and Sasaki, 1977
Silk worm pupal oocyte	25µg	-	Mutagenicity not observed	Kawachi <i>et al.</i> , 1980 and Tazima, 1980
Male rat	300mg/kg bw orally	-	Mutagenicity not observed	Tanaka <i>et al.</i> , 1980
Mouse, hamster, rat hepatocytes	-	-	DNA repair found	McQueen et al.,1981
Rat	12000, 3000,6000, ppm	-	Mutagenicity not seen	Haworth et al., 1981
Mice	420, 61 mg/kg BW orally	25 h	SCE rate rise	Parodi et al., 1982,1983
Mice	30, 0, 3, 10 mg/kg BW in water	103 no. of weeks	Carcinomas of hepatic cells and spleen	Chhabra <i>et al</i> , 1990
-	-	-	Carcinomas and sarcomas risk	Ingle <i>et al.</i> , 2023

Table-6 Study of aniline on genes and cancer

3.6 Study of aniline on thyroid gland- The study conducted by [48] For five days, two weeks, four weeks, and thirteen

weeks, female F344 rats were given nutritional feed containing 0, 50, 200, 375, 500, or 750 parts per million of 4,4'-methylenebis(N'-dimethyl) aniline (MDA). During study weeks 6 through 13, the 750ppm group's mean body weight

dropped by 5% in comparison to the control. With increasing meal concentrations of MDA and exposure duration, blood TSH rose while serum T4 and T3 levels fell.

Rats given aniline had thyroid gland histopathology that revealed more micro follicles, less colloid secretion, vacuolation in the colloid, a breakdown of the capsular layer, follicular disruption, thyroid follicle fusion, and more interfollicular space than the control group. Rats treated with aniline had significantly lower levels of the thyroid hormones triiodothyronine (T3) and tetraiodothyronine (T4) and higher levels of thyroid stimulating hormone (TSH) when their thyroid glands were analysed hormonal. Upon these findings, it can be said that aniline may have a negative impact on the thyroid gland's structure and function [49].

Animal model	Aniline dose	Time period	Inference	Reference
Rats	750,0,	13 WK, 4 WK, 5	Follicular cell hyperplasia in thyroid gland. TSH elevate	Dodd et al.,
	50,200,375,500ppm	days, 2 WK	and serum T4 and T3 become less with increasing MDA	2012.
		duration	with exposure time	
Rat	-	-	Thyroid follicle fusion, follicular disruption, dissolution	Dhurvey et
			of the capsular layer, vacuolation in the colloid, decrease	al., 2021.
			in colloid secretion, and an increase in micro-follicles	
			interfollicular space, elevated thyroid stimulating	
			hormone (TSH) levels, and decreased levels of the	
			thyroid hormones triiodothyronine (T3) and	
			tetraiodothyronine (T4).	

Table- 7 Study of aniline on thyroid gland

3.7 Study of aniline on respiratory system- Aniline's main effects on the human lung, such as upper respiratory tract irritation and congestion, are both acute (short-term) and chronic (long-term) [50].

Aniline inhalation can irritate the respiratory tract, resulting in coughing or breathing difficulties [51].

Table-8 Study of aniline on respiratory system

Animal model	Aniline dose	Time period	Inference	Reference
Humans	-	-	Upper respiratory tract irritation and congestion	US Environment protection agency, 2000.
Humans	-	-	Irritation of respiratory tract cough along with breathing difficulties	NIH and Feng et al., 2020.

3.8 Study of aniline on endocrine system- It was shown by [52] aniline (aminobenzene) given subcutaneously to rats causes a significant decrease in the blood corticosterone level with a marked enlargement and ivory-white discoloration of the adrenal glands. The sensitivity of the adrenals to stressful stimuli (e.g. vasopressin and insulin) and to exogenous ACTH is also considerably reduced; thus, it seems reasonable to assume that aniline inhibits the production of corticoids by

primarily affecting the adrenocortical cells. The lack of circulating corticoids leads to a compensatory hypersecretion of ACTH, resulting in adrenocortical hyperplasia.

The study used the human adrenal H295R cell line and adult male Danio rerio zebrafish. Tested aniline derivatives all

decreased the H295R cell line's testosterone (T) levels. The disruption of sex hormones could be attributed to regulatory alterations in numerous steroidogenic genes, including the decrease in number of gene-regulation of StAR or CYP17 factors and the increase in number of receptors of CYP19A, that was seen in the H295R cells. T levels decreased and E2/T ratios increased in male zebrafish, which showed essentially similar patterns of change. Again, the fish's sex hormone changes might be explained by a down-regulation of key steroidogenic genes such as cyp17 or 3 β -hsd, but a slight up-regulation of the cyp19a gene [54].

In a study by [53], aniline at a concentration of 10.0 g/ml was applied for 48 hours to NCI-H295R adrenocortical human cells. Endocrine disruption was found to have anti-prostaglandin and anti-androgenic effects.

Animal model	Aniline dose	Time	Inference	Reference
Rats	-	- -	Decrease in the blood corticosterone level with a marked enlargement and ivory-white discoloration of the adrenal glands.	Kovacs <i>et</i> <i>al.</i> ,1971
NCI-H295R adrenocortical human cells	Concentration of 10.0g/ml	48 hours	Endocrine disruption was observed as anti-androgenic an anti-prostaglandin effect.	Albert <i>et al.</i> , 2013
Adult male zebrafish and Human adrenal H295R cell line	-	-	Decreased testosterone (T) levels. down- regulation of key steroidogenic genes like cyp17 or 3β -hsd, although the fish's modest up-regulation of the cyp19a gene may account for the sex hormone changes.	Bhuiyan <i>et</i> <i>al.</i> , 2019

 Table-9 Study of aniline on endocrine system

3.9 Study of aniline on blood- A study carried out by [56] A 6-hour exposure to 2 ppm aniline resulted in a time-dependent increase in blood Met-Hb in 19 nonsmokers. The highest Met-Hb level in the blood was measured (mean 1.21 ± 0.29 percent, range 0.80–2.07%). Over the course of 24 hours, the mean Met-Hb level (0.65 ± 0.18 %) returned to the baseline level (0.72 ± 0.19 %).

A 6-hour inhalation trial involving exposure to 2 ml aniline/m3 revealed elevated methaemoglobin levels in both healthy male and female subjects. From the baseline level of 0.7% to 1.2% methaemoglobin, the median methaemoglobin augmentation was 0.5% [57].

After study by [58] evaluated the aniline's 96-hour LC50, two sublethal concentrations of aniline (4.19 mg/l and 8.39 mg/l) were selected for acute exposure testing in freshwater feeder fish Channa punctatus. Blood was drawn 24, 48, 72, and 96 hours after exposure to examine the genotoxic, cytotoxic, and haematological effects of sublethal dosages of aniline in C. punctatus. Time and dose-dependent DNA damage increased significantly in the comet and micronuclei assays, with the most damage happening 96 hours after exposure. Analyses using ATR-FTIR and scanning electron microscopy after aniline exposure showed structural anomalies and modifications in the biomolecules of the red blood cells in the group exposed to aniline as compared to the control group, respectively.

In one investigation by [55], thirty male Wistar rats per group received nose-only treatment to mean analytical values of 9.2, 32.4, 96.5, and 274.9 mg aniline/m3. Six hours a day, five days a week, were administered to the rats for two weeks

(days 0–11), after which they were given two weeks (day 28) to recuperate. Serial sacrifices were made for certain experiments on days 28, 14, 11, and 4. The primary causes of toxicity, erythrocytotoxicity and haemoglobin synthesis, were among the changes observed.

A study by [54] investigated the histological, biochemical, and haematological responses of rats to sub chronic exposure to aniline hydrochloride. The male Sprague-Dawley rats were given 600 ppm of AH in their drinking water, while the control rats were given tap water only. At 30, 60, and 90days following treatment, five rats per group were killed. The spleen organ-to-body weight ratio of the AH-treated rats was 56, 61, and 53% higher than that of the controls at days 30, 60, and 90, respectively.

Animal	Aniline dose	Time period	Inference	Reference
model				
Male albino	600 ppm	90, 30, 60 days	In the AH-treated rats, the spleen's organ-to-	Khan et
rat			body weight ratio was 56, 61, and 53% greater.	al., 1993.
Male wistar	274.9, 9.2,	6 hours per day, five days per	Methemoglobin formation and erythrocyte	Pauluhn,
rats	96.5, 32.4 mg	week, for two weeks (days	toxicity.	2004.
	aniline/m ³	0-11), and then two weeks		
		after exposure (day 28).		
Human	2 ppm	6-h exposure	An increase in urine aniline and blood Met-Hb	Kafferlein et
				al., 2014.
Fish	8.39 mg/l and	96, 24, 72, 48 h	Studies using ATR-FTIR and scanning electron	Sharma et
Channa	4.19 mg/l)		microscopy revealed structural irregularities	al., 2023.
punctatus			and changes in the biomolecules of RBCs from	
			the group exposed to aniline.	

Table- 10 Study of aniline on blood

4. Conclusion

Based on available information, aniline and its derivatives alter the biological behaviour of various organisms. Higher of aniline cause the blood doses to produce methemoglobinemia and excessive reactive oxygen species. It also increases the risk of developing carcinomas and sarcomas. The reproductive systems of both sexes are affected. Aniline reduces the fecundity of the animal. This is mostly due to excessive oxidative stress caused by aniline. Thus, we conclude that aniline and its derivatives are detrimental to biological systems and they should be used with utmost care. Hence, it is important to derive safety guidelines and search for alternative natural substances to use in place of aniline. There is scope for future researchers to study about the mechanisms and precise effects of aniline and it's derivative's toxicity on living organisms.

Author contributions

The authors NK and VD conceived the idea, and prepared the protocol. NK did the literature search and that was counterchecked by VD. NK prepared the 1st draft of the manuscript and VD critically edited the article. All of them agreed to the final manuscript.

Data availability statement

The research articles used in this systematic review are available for use. Ethical approval Institutional review board approval is not required.

Declaration of patient consent

Patient's consent not required as there are no patients in this study.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

Use of artificial intelligence (AI)-assisted technology for manuscript preparation

The authors confirm that there was no use of artificial intelligence (AI)-assisted technology for assisting in the writing or editing of the manuscript and no images were manipulated using AI

Acknowledgements

We are grateful to our library colleagues for their assistance with the literature. We are also grateful to our university for their initiatives to support and foster research. We are also grateful for the constant support of Dr. V. T. Dhurvey and Miss. Vaibhavi Ingle.

References

- M. Anjalina, N. Kanagathara and R. Suganthi, "A brief review on aniline and its derivatives," *Materials Today:* proceedings, Vol. 33, Issue. 6, pp. 4751-4756, 2020.
- [2] R. J. Lewis, "Hawley's condensed chemical dictionary 15th edition" New York, Vol. 1, pp. 385, 2007.
- [3] M. Mattarozzi, F. Lambertini, M. Suman M. Careri "Liquid chromatography full scan- high resolution mass spectrometry-based method towards the comprehensive analysis of migration of primary aromatic amines from food packaging," *Journal of Chromatography A*, Vol.1320, pp. 96-104, 2013.
- [4] A. Saleh, S. Molaei, F. Sheijooni, E. Abedi "Antifouling paint booster biocides (Irgarol 1051 and diuron) in marinas and ports of Bushehr," *Marine Pollution Bulletin*, Vol.105, Issue.1, pp.367-373, 2016.
- [5] Y. Wang, H. Gao, X. Lin, S. Dong, H. Dong, et al., "Aniline induced oxidative stress and apoptosis of primary cultured hepatocytes," *International Journal of Environmental Reasarch and Public Health*, Vol.12, Issue 2, pp. 1188, 2016.
- [6] EC."Recommendation for substance 3,4-dichloroaniline" http://echa.europa.eu/information-onchemicals/ information from existing substances regulation, Vol. 3, pp. 201. 2016.
- [7] F. Di Girolamo, L. Campanella, R. Samperi, A. Bachi, "Mass spectrometric identification of hemoglobin modifications induced by nitroso benzene," *Ecotoxicology and Environmental Safety*, Vol. 72, Issue 6, pp. 1601-1608, 2009.
- [8] M. Sihtmae, M. Mortimer, A. Kahru, I. Blinova, "Toxicity of five anilines to crustaceans, protozoa and bacteria," *Journal* of Chemical Sciences, Vol. 75, Issue 6, pp. 1291-1298, 2010.
- [9] M. Akyuz, S. Ata, "Determination of aromatic amines in hair dye and henna samples by ion-pair extraction and gas chromatography-mass spectrometry," *Journal of Pharmaceutical and Biomedical Analysis*, Vol. 47, Issue 1, pp. 68-72, 2008.
- [10] D. Pezo, M. Fedeli, O. Bosetti, C. Nerin, "Aromatic amines from polyurethane adhesives in food packaging: the challenge of identification and pattern recognition using Quadrupole-Time of Flight-Mass Spectrometry," *Analytica Chimica Acta*, Vol. **756**, pp. **31- 49**, **2012**.
- [11] T. Kawakami, K. Isama, H. Nakashima, T. Tsuchiya, A. Matsuoka, "Analysis of primary aromatic amines originated from azo dyes in commercial textile products in Japan," *Journal Environmental Science and Health*, Vol. 45, pp. 1281-1288, 2010.
- [12] V. Marlatt, P. Lopez Martínez, J. Lopez de Alba, V. Castrejon Duran, "Optical fibre reflectance sensor coupled to a multistring flow injection system for preconcentration and determination of 1-naphthylamine in water samples," *Analytica Chimica Acta*, Vol. 574, Issue. 2, pp. 406-411. 2006.
- [13] C. J. Martyniuk, "Biological responses to phenylurea herbicides in fish and amphibians' new directions for characterizing mechanisms of toxicity," *Environmental Toxicology and Pharmacology*, Vol. 34, pp.9-18, 2017.
- [14] G. Campenella, M. Ghaani, G. Quetti S. Farris, "On tsshe origin of primary aromatic amines in food packaging materials," *Trends in Food Science and Technology*, Vol. 3, pp. 1-8, 2015.
- [15] B. Jurado-Sanchez, E. Ballesteros, M. Gallego, "Occurrence of aromatic amines and N-nitrosamines in the different steps

of a drinking water treatment plant," *Water Research*, Vol. 46, Issue 11, pp. 4543-4553, 2012.

- [16] S. Boulahlib, A. Boudina, K. Si-Ahmed, Y. Bessekhouad, M. Trari, "Development and validation of a fast and simple HPLC method for the simultaneous determination of aniline and its degradation products in waste water," *Journal Analytical Chemistry*, Vol. 8, Issue 9, pp. 5949-5956, 2016.
- [17] US Geological Survey. "Analysis of the herbicide diuron, three diuron degrades and six neonicotinoid insecticides in water-method details and application to two georgia streams," *scientific investigations report*, pp 2012-5206, 2012.
- [18] M. Akyuz S. Ata, "Simultaneous determination of aliphatic and aromatic amines in water and sediment samples by ionpair extraction and gas chromatography-massspectrometry," *J ournl Chromatography A*, Vol. **1129**, Issue. **2**, pp **88-94**, **2006**.
- [19] N. Kim, W. Shim, U. Yim, S. Hong, S. Ha, G. Han, K. Shin, "Assessment of TBT and organic booster biocide contamination in seawater from coastal areas of South Korea," *Marine Pollution Bulletin*, Vol.**78**, Issue. **1-2**, pp. **201-208**, **2014**.
- [20] Y. Okazaki, K. Yamashita, M. Sudo, M. Tschitani, I. Narama, R. Yamaguchi, et al., "Neurotoxicity induced by a single oral dose of aniline in rats," *Journal of Veterinary Medical Science*, Vol. 63, pp.539, 2001
- [21] P. Makhdoumi, H. Hossini, G. Ashraf, M. Limoee, "Molecular mechanism of aniline induced spleen toxicity and neuron toxicity in experimental rat exposure," *Current Neuropharmacology*, Vol.17, Issue. 3, pp. 201-214, 2019.
- [22] G. Rankin, D. Yang, K. Cressey-Veneziano, S. Casto, R. Wang, et al., "In vivo and in vitro nephrotoxicity of aniline and its monochlorophenyl derivatives in the fischer 344 rat," *Toxicology*, Vol. 38, Issue. 3, pp. 269-283,1986.
- [23] M. Yoshida, H. Yoshikawa, H. Goto, I. Hara I, "Evaluation of the nephrotoxicity of aromatic nitro-amino compounds by urinary enzyme activities," *Journal Toxicological Sciences*, Vol.14, Issue. 3, pp.257-265, 1989.
- [24] S. Hong, D. Anestis, T. Henderson, G. Rankin, "Haloanilineinduced in vitro nephrotoxicity effects of 4-haloanilines and 3,5-dihaloanilines," *Toxicology Letters*, Vol.114, pp.125, 2000.
- [25] S. Ramteke V. Dhurvey "Ameliorating action of astragalus membranous on kidney disease of male rat," *International Journal of Current Research in Science and Engineering*, Vol. 11, pp. 2393-2398, 2020.
- [26] F. Jenkins, J. Robinson, J. Gellatly G. Salmond, "The noeffect dose of aniline in human subjects and a comparison of aniline toxicity in man and the rat," *Food Cosmetics Toxicology*, Vol. 5, Issue.5, pp.671-679. 1972.
- [27] P. Eyer, H. Kampffmeyer, H. Maister, "Biotransformation of nitrobenzene, phenylhydroxylamine and aniline in the isolated perfused rat liver," Xenobiotica, *The fate of foreign compound biological system*, Vol. 7, Issue.7, pp. 499-511, 1980.
- [28] R. Chhabra "Carcinogenicity of P-chloroaniline in rats and mice," *Food and Chemical Toxicology*, Vol. 2, pp 119-125, 1991.
- [29] S. Sharma, G. Goyal, A. Sharma, "Toxicity of tomato red, a popular food dye blend on male albino mice," *Toxicologic pathology*, Vol. 6, pp. 51-58, 2008.
- [30] C. Koenig, C. Beevers, K. Pant, "Assessment of the mutagenic potential of para- chloro-aniline and aniline in the

liver, spleen, and bone marrow of big blue rats with micronuclei analysis in peripheral blood," *Environmental and Molecular Mutagenesis*, Vol. 9, pp. 785-794, 2018.

- [31] P. Mohurle, V. Dhurvey R. Urkude, H. Pawar, D. Joshi, "A mini review on effect of aniline on liver and spleen," *Journal of Environmental Science Research and Technology*, Vol.12, pp.28-35, 2023.
- [32] S. Hatakeyama, K. Kovacs, E. Yeghiayan, J. Blascheck, "Aniline induced changes in the corpora lutea of rats," *American Journal of Obstetrics and Gynecology*, Vol. 311, pp. 400-405, 1971.
- [33] C. Price, R. Tyl, T. Marks, L. Paschke T. Ledoux, J. Reel, "Teratologic and postnatal evaluation of aniline 344 hydrochloride in the Fischer rat," Toxicology Applied Pharmacology, Vol. 3, pp. 465-474, 1985.
- [34] R. Nair, C. Auletta, R. Schroder, F. Johannsen, "Chronic toxicity, oncogenic potential, and reproductive toxicity of p-Nitroaniline in rats," *Toxicological Sciences*, Vol. 3, pp. 607-616, 1990.
- [35] A. Sardo, U. Azeiteiro, U. Pereira L. Morgado A. Soares, "Histological evaluation of the exposure to 3,4dichloroaniline in the estuarine mysid Mesopodopsis slabberi, under experimental conditions" *Fresenius Environmental Bulletin*, Vol.7, pp. 579-586, 2005.
- [36] B. Zhang, S. Lin, "Effects of 3,4-Dichloroaniline on testicle enzymes as biological markers in rats," *Journal of Biomedical Research*, Vol. 1, pp. 40-47, 2009.
- [37] F. Eissa, E. Makawy, "Assessment of 3,4-Dichloroaniline toxicity as environmental pollutant in male mice," *Egyptian Journal of Basic and Applied Sciences*, Vol.3, Issue 2, pp. 73-80, 2012.
- [38] J. Holm, S. Mazaud-Guittot, N. Danneskiold-Samsoe, C. Chalmey, B. Jensen, M. Norregard, "Intrauterine exposure to paracetamol and aniline impairs female reproductive development by reducing follicle reserves and fertility," *Toxicological Sciences*, Vol. 1, pp. 178-187, 2016.
- [39] M. Bhuiyan, "Effects of aniline and several aniline derivatives on sex hormone regulation and reproduction of adult Zebrafish," Ph.D Thesis. Seoul National University, 2019.
- 40] M. Ibrahim, S. Zulkifli, M. Azmai, F. Mohamat-Yusuff, A. Ismail "Reproductive toxicity of 3,4-dichloroaniline (3,4-DCA) on javanese medaka," *Animals*, Vol.11, Issue 3, pp. 789, 2021.
- [41] V. Ingle, V. Dhurvey, R. Urkude "Genotoxic, carcinogenic and reproductive studies on aniline a mini review," *International Journal of Scientific Research in Biological Sciences*, Vol. 10, pp. 42-51, 2023.
- [42] S. Abe, M. Sasaki, "Chromosome aberrations and sister chromatid exchanges in Chinese hamster cells exposed to various chemicals," *Journal of the National Cancer Institute*, Vol.6, pp. 1635-1644. 1977.
- [43] T. Kawachi, T. Yahagi, T. Kada, Y. Tazima, M. Ishidate, M. Sasaki, "Cooperative programme on short-term assays for carcinogenicity in Japan," *IARC Scientific Publications*, Vol. 2, pp. 323-330, 1980.
- [44] K. Tanaka, S. Marui, T. Mii, "Mutagenicity of extracts of urine from rats treated with aromatic amines," *Mutation Research Genetic Toxicology and Environmental Mutagenesis*, Vol. 2, pp. 173-178, 1980.
- [45] C. Mcqueen, C. Maslansky, B. Crescenzi, G. Williams "The genotoxicity of 4,4'- Methylenebis-2-chloroaniline in rat,"

Toxicology and Appllied Pharmacology, Vol.9, 231-237, 1981.

- [46] S. Parodi M. Taningher, P. Russo, M. Pala, M. Tamaro, C. Monti Bragadin, "DNA- damaging activity in vivo and bacterial mutagenicity of sixteen aromatic amines and azoderivatives, as related quantitatively to their carcinogenicity," *Carcinogenesis*, Vol. 2, Issue. 11, pp.1317-1326, 1981.
- [47] R. Chhabra, "Carcinogenicity of P-chloroaniline in rats and mice," *Food and Chemical Toxicology*, Vol. 2, pp. 199-125, 1991.
- [48] D. Dodd, L. Pluta, M. Sochaski M K. Funk R. Thomas "Subchronic thyroid toxicity evaluation of 4,4'-methylenebis (N, N'-dimethyl) aniline in Fischer 344 rats," *Journal of Toxicology and Environmental Health Sciences*, Vol.11, pp. 637-644, 2012.
- [49] V. Dhurvey, N. Kumbhare, F. Karim, S. Katke, "Aniline induced histological alterations and evaluation of serum hormones in thyroid gland of male albino rats," *Advances in Geriatric and Pediatric Endocrinology*, Vol. 2, pp. 31-38. 2021.
- [50] Environmental protection agency united states, 2000.
- [51] The National Institutes of Health NIH, 2020.
- [52] K. Kovacs, J. Blascheck, E. Yeghiayan S. Hatakeyama, C. Gardell, "Adrenocortical lipid hyperplasia induced in rats by aniline," A histologic and electron microscopic study. *American Journal of Pathology*, Vol. 1, pp.17-34, 1971.
- [53] O. Albert, C. Desdoits-Lethimonier, L. Lesne, "Paracetamol, aspirin and indomethacin display endocrine disrupting properties in the adult human testis in vitro" *Human Reproduction*, Vol. 28, Issue. 7, pp.1890-1898, 2013.
- [54] F. Khan, S. Kaphalia, J. Boor, S. Ansari, "Subchronic toxicity of aniline hydrochloride in rats," *Archives of Environmental Contamination and Toxicology*, vol. 24, Issue. 3, pp. 368-374, 1993.
- [55] J. Pauluhn, "Subacute Inhalation toxicity of aniline in rats' analysis of time-dependence and concentration-dependence of hematotoxin and splenic effects," *Toxicological Sciences*, Vol. 81, Issue.1, pp.198-206, 2004.
- [56] H. Kafferlein, H. Broding, J. Bunger B. Jettkant, S. Koslitz, M. Lehnert, "Human exposure to airborne aniline and formation of methemoglobin a contribution to occupational exposure limits," *Archives Toxicology*, Vol.88, Issue. 9, pp.1419-1431, 2014.
- [57] MKA value documentation, 2018.
- [58] G. Sharma, P. Chadha "Evaluation of hematological, genotoxic, cytotoxic and ATR- FTIR alterations in blood cells of fish Channa punctatus after acute exposure of aniline. *Scientific reports* Vol. 67, Issue. 1, pp.213, 2023.

AUTHOR'S PROFILE

Mr. Nayan Kumbhare He has finished his B. Sc and M.Sc from RTM Nagpur University, Nagpur in the year 2017 and 2019 respectively. He is currently working as a research scholar in RTM Nagpur University, Nagpur. The topic of his research is 'Effect of aniline on testis epididymis and sperm



analysis of male albino rat'. He has qualified MH- SET and GATE exam. He has a teaching experience of 2 years and a research experience of 2 years. He has also published 2 research papers in international journals.

Dr. Varsha Dhurvey She has finished her M.Sc and Ph.D. form RTM Nagpur University, Nagpur in 1993 and 2003 respectively. She is currently working as a professor and Head in the Dept. of Zoology, RTM Nagpur University, Nagpur. She has a teaching and research experience of 22 years. She



has published more than 55 research papers in reputed national and international journals, Thomson Bruter (ICI and Web of science). Her main research work focuses on reproductive physiology and endocrinology.